

## AQ97 HiFi Hot Start 2x Master Mix RED

2.0 mM MgCl<sub>2</sub> final concentration



Cat. No.: A810801

A810801

100 Reactions

MADE IN DENMARK

| AQ97 HiFi Hot Start 2x Master Mix RED |             |
|---------------------------------------|-------------|
| ID No.                                | 5500670     |
| Cap colour                            | Clear       |
| Content                               | 2 x 1.25 ml |

### Contents

All-in-one 2x master mix containing the AQ97 High Fidelity Hot Start DNA Polymerase, optimized buffer, dNTPs, MgCl<sub>2</sub> and inert red dye for direct loading on agarose gel. Recommended for low-bias, high fidelity amplification.

There is no need for additional loading dyes. Simply load a portion of the reaction product onto an agarose gel. The red dye front runs at 1000 – 2000 bp on a 0.5 – 1.5 % agarose gel.

### Recommended Storage and Stability

| Temperature      | Duration                         |
|------------------|----------------------------------|
| Room temperature | Up to 5 days                     |
| 4 °C             | Up to 6 months                   |
| -20 °C           | Long term.<br>See expiry on tube |

### Protocol

- AQ97 HiFi Hot Start 2x Master Mix RED is not frozen at -20 °C. Simply mix the tube thoroughly and it is ready for preparation of the reaction mix.

1. Combine master mix, primers, template DNA and water according to the following table.

**Table 1:** Recommended reaction setup

| Component                  | Vol./reaction*  | Final concentration*  |
|----------------------------|-----------------|---|
| 2x Master Mix              | 25 µl           | 1x  |
| Primer A (10 µM)           | 1 µl            | 0.2 µM  |
| Primer B (10 µM)           | 1 µl            | 0.2 µM  |
| 25 mM MgCl <sub>2</sub>    | 0 µl (0 – 5 µl) | 2.0 mM (2.0 – 4.5 mM)   |
| Betaine (5 M)**            | 0 – 5 µl        | 0 – 0.5 M   |
| PCR-grade H <sub>2</sub> O | X µl            | -   |
| Template DNA               | X µl            | genomic DNA: 50 ng (10 – 500 ng)<br>plasmid DNA: 0.5 ng (0.1 – 1 ng)<br>bacterial DNA: 5 ng (1 – 10 ng) |
| <b>TOTAL volume</b>        | 50 µl           | -   |

\* Suggested starting conditions; theoretically used conditions in brackets.

\*\* Suggested for GC-rich amplification. See section *Strategies for Optimization*.

2. Transfer the appropriate volume to a PCR plate or strip compatible with the chosen thermal cycler. Seal the plate/strip.

### 3. Three-step PCR program

**Table 2:** Recommended cycling conditions

| Step                 | Duration of cycle   | Temperature                  |
|----------------------|---|------------------------------|
| Initial denaturation | 30 sec – 2 min  | 98 °C                        |
| 25 – 35 cycles       | 10 – 20 sec <sup>a)</sup><br>15 – 30 sec <sup>b)</sup><br>10 – 60 sec <sup>c)</sup> | 98 °C<br>55 – 70 °C<br>72 °C |
| Final elongation     | 5 min   | 72 °C                        |

<sup>a</sup>. Denaturation: During thermocycling, 10 seconds usually works very well. Longer denaturation times might be required for long range PCR or amplification from templates with a high GC content.

<sup>b</sup>. Primer annealing: Typically, the annealing temperature is about 3 – 5 °C below the T<sub>m</sub> (melting temperature) of the primers used. **Because of the high salt content within the AQ97 HiFi Hot Start 2x Master Mix RED, the annealing temperature will likely be higher than with more traditional PCR buffers.**

<sup>c</sup>. Extension: The recommended extension temperature is 72°C. Extension times highly depends on the length of the amplicon. **Generally, we recommend an extension time of 10-30 seconds per kb for complex genomic targets. 10 seconds per kb is often sufficient for simpler targets (such as plasmids) or short complex targets (< 3 kb). 30-60 seconds per kb is recommended for long amplicons (> 3 kb).**

### 4. Two-step PCR program

For targets with annealing temperatures ≥ 72°C (usually GC-rich primers), a 2-step thermocycling protocol (combining annealing and extension into one step) can be used.

## Strategies for Optimization

#### High yield/Nonspecific amplification

- Decreased primer concentration can improve PCR product specificity of long amplicons.
- Decreased extension time can improve PCR product specificity of short amplicons.
- Decreased PCR cycles will decrease PCR product yield.

#### Long-range amplification

- Longer extension times often resolve low-yield amplification of long amplicons.
- Increased template concentration will increase product yield.
- Increased number of PCR cycles will increase product yield.
- Increased primer concentration can increase product yield for some reactions.

#### GC-rich amplification

- The addition of 0.5 M Betaine solution often improves reaction performance.
- Primers of 20 – 40 nucleotides with a GC content of 40 - 60 % are recommended. Online Software such as the [Primer3plus](#) can be used to design primers.

#### MgCl<sub>2</sub>

- The optimal MgCl<sub>2</sub> concentration should be determined empirically but in most cases a concentration of 2.0 mM, as provided in this Master Mix (1x), will produce satisfactory results.

For Research Use Only. Not for use in diagnostics procedures.

Other product sizes, combinations and customized solutions are available. Please look at [www.ampliqon.com](#) or ask for our complete product list for PCR Enzymes. For customized solutions please contact us.

**Made in Denmark**

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