

Taq DNA Polymerase 1.1x Master Mix

2 mM MgCl₂ final concentration



Cat. No.: A130303

A130303

Cat. No.	Size Reactions	Taq DNA Polymerase 1.1x Master Mix, 2 mM MgCl ₂
ID No.	-	5200050
Cap colour	-	Blue
A130303	500	15 x 1.5 ml

Key Features

- High performance thermostable DNA polymerase
- Significantly reduced set up time
- Diminished risk of contamination
- Increased reproducibility
- No proofreading – lacks a 3'→5' exonuclease activity
- Ideal for TA cloning – leaves an A' overhang

Taq DNA Polymerase 1.1x Master Mix is a ready-to-use 1.1x reaction mix with the Ampliqon Taq DNA polymerase, the NH₄⁺ buffer system, dNTPs and magnesium chloride present. Each reaction requires 45 µl of the 1.1x Master Mix. Simply add primers, template and water to a total reaction volume of 50 µl to successfully carry out primer extensions and other molecular biology applications.

Taq DNA Polymerase 1.1x Master Mix offers several advantages. Set up time is significantly reduced. The chance of contaminating component stocks is eliminated. Reduction of reagent handling steps leads to better reproducibility. Standard tests can be set up with the confidence that results will be consistent every time.

Composition of the Taq DNA Polymerase 1.1x Master Mix (2 mM MgCl₂ final concentration)

- Tris-HCl pH 8.5, (NH₄)₂SO₄, 2.2 mM MgCl₂, 0.11% Tween[®] 20
- 0.22 mM of each dNTP
- 0.11 units/µl Ampliqon Taq DNA polymerase
- Stabilizer

Recommended Storage and Stability

Long term storage at -20 °C. Product expiry at -20 °C is stated on the label.

Option: Store at +4 °C for up to 6 months.

Quality Control

Taq DNA Polymerase is tested for contaminating activities, with no traces of endonuclease activity, nicking activity or exonuclease activity.

Unit Definition

One unit is defined as the amount of polymerase that incorporates 10 nmoles of dNTPs into acid-precipitable DNA in 30 minutes at 72 °C under standard assay conditions.

Protocol

This protocol serves as a guideline for primer extensions. Optimal reaction conditions such as incubation times, temperatures, and amount of template DNA may vary and must be determined individually.

Note:

- Set up reaction mixtures in an area separate from that used for DNA preparation or product analysis. Work on ice at all times.

1. Thaw Taq 1.1x Master Mix and primers. **It is important to thaw the solutions completely and mix thoroughly before use to avoid localized concentrations of salts.** Keep all components on ice.
2. Set up each reaction. Table 1 shows the reaction set up for a final volume of 50 µl. If desired, the reaction size may be scaled down. Use 18 µl of the Taq 1.1x Master Mix in a final volume of 20 µl.
If your template DNA is very dilute, it might be an advantage to use the Ampliqon Taq 2x Master Mix. (See Additional Products)

Table 1. Reaction components (reaction mix and template DNA)

Component	Vol./reaction	Final concentration
Taq 1.1x Master Mix	45 µl	1x
Primer A	1 µl (10 µM)	0.2 µM (0.1 – 1.0 µM)
Primer B	1 µl (10 µM)	0.2 µM (0.1 – 1.0 µM)
PCR-grade H ₂ O	X µl	-
Template DNA	X µl	genomic DNA: 50 ng (10 – 500 ng) plasmid DNA: 0.5 ng (0.1 – 1 ng) bacterial DNA: 5 ng (1 – 10 ng)
TOTAL volume	50 µl	-

3. Mix the reaction mix thoroughly and dispense appropriate volumes into reaction tubes. Mix gently, e.g. by pipetting the reaction mix up and down a few times.
4. Add template DNA to the individual tubes containing the master mix.
5. Program the thermal cycler according to the manufacturer's instructions.
For maximum yield and specificity, temperatures and cycling times should be optimized for each new template target or primer pair.
6. Place the tubes in the thermal cycler and start the reaction.

Three-step PCR program

Cycles	Duration of cycle	Temperature
1	2 – 5 minutes	95 °C
25 - 35	20 – 30 seconds ^a 20 – 40 seconds ^b 30 seconds ^c	95 °C 50 – 65 °C 72 °C
1	5 minutes ^d	72 °C

- ^a Denaturation step: This step is the first regular cycling event and consists of heating the reaction to 95 °C for 20 – 30 seconds. It causes melting of the DNA template by disrupting the hydrogen bonds between complementary bases, yielding single-stranded DNA molecules.
- ^b Annealing step: The reaction temperature is lowered to 50 – 65 °C for 20 – 40 seconds allowing annealing of the primers to the single-stranded DNA template. Typically, the annealing temperature is about 3 – 5 °C below the T_m of the primers used.
- ^c Extension/elongation step: Taq polymerase has its optimum activity temperature at 72 °C. At this step the DNA polymerase synthesizes a new DNA strand complementary to the DNA template strand. The extension time depends on the length of the DNA fragment to be amplified. As a rule of thumb, at its optimum temperature the DNA polymerase will polymerize a thousand bases per minute.
- ^d Final elongation: This single step is occasionally performed at a temperature of 72 °C for 5 minutes after the last PCR cycle to ensure that any remaining single-stranded DNA is fully extended.

Related Products

Taq Polymerase (500 units) *	Cat. No.
Taq DNA Polymerase 5 U/μl • with 10x Ammonium Buffer • 5x PCR Buffer RED	A110003 A111103 A111803
Taq DNA Polymerase 5 U/μl, RED • with 10x Ammonium Buffer	A200003 A201103
Taq DNA Polymerase 5 U/μl, glycerol free • with 10x Ammonium Buffer	A100003 A101103

Hot Start Polymerase (500 units) *	Cat. No.
TEMPase Hot Start DNA Polymerase, 5 U/μl • with 10x Ammonium Buffer • 5x PCR Buffer RED	A220003 A221103 A221803
TEMPase Hot Start DNA Polymerase, glycerol free 5 U/μl • with 10x Ammonium Buffer	A240003 A241103

High Fidelity - Proof reading (500 units) **	Cat. No.
AccuPOL DNA Polymerase 2.5 U/μl • with 10x Ammonium Buffer	A210003 A211103

*Available in kits including one or two buffers (Ammonium Buffer, Standard Buffer or Combination Buffer). **AccuPOL only available in kits with Ammonium Buffer. All kits include extra 25 mM MgCl₂.

Buffers for DNA polymerases *	Cat. No.
10x Ammonium Buffer, 3 x 1.5 ml	A301103
10x Standard Buffer, 3 x 1.5 ml	A302103
10x Combination Buffer, 3 x 1.5 ml	A303103
5x PCR Buffer RED, 6 x 1,5 ml **	A301810

*Ammonium Buffer, Standard Buffer and Combination Buffer are also available as Mg²⁺ free buffers, detergent free buffers and Mg²⁺ and detergent free buffers. **For direct gel loading and visualisation.

Taq Master Mixes (500 x 50 μl reactions) *	Cat. No.
2x Master Mix, 1.5 mM MgCl ₂ final concentration	A140303
2x Master Mix RED, 1.5 mM MgCl ₂ final concentration	A180303

TEMPase Hot Start Master Mixes (500 x 50 μl reactions) *	Cat. No.
2x Master Mix A**, 1.5 mM MgCl ₂ final concentration	A230303
2x Master Mix A**BLUE, 1.5 mM MgCl ₂ final concentration	A290403

*Master mixes available also in 1.1x variants as well as 2 mM MgCl₂ variants, **Mix A is Ammonium Buffer based, also available as Mix C based on Combination Buffer.

Special Master Mixes (500 x 50 μl reactions)	Cat. No.
Multiplex 2x Master Mix, 3 mM MgCl ₂ final concentration	A260303
GC TEMPase 2x Master Mix I – for GC-rich templates	A331703
GC TEMPase 2x Master Mix II – for GC-rich templates	A332703

Real-time PCR Master Mixes (400 x 25 μl reactions)	Cat. No.
RealQ Plus 2x Master Mix for probe, • without ROX™ • with low ROX™ • with high ROX™	A313402 A314402 A315402
RealQ Plus 2x Master Mix Green • without ROX™ • with low ROX™ • with high ROX™	A323402 A324402 A325402

Ultrapure dNTPs*	Cat. No.
dNTP Mix 40 mM (2 x 500 μl): 10 mM each dA, dC, dG, dT	A502004
dNTP Set, 100 mM each: 250 μl of each dA, dC, dG and dT	A511104

*Other concentrations and Single dNTPs are available.

Loading Buffers and Ladders	Cat. No.
5x Loading Buffer Red *, 5 x 1 ml	A608104
PCR DNA Ladder **, 100 – 3000 bp, 1 x 0.5 ml	A610341

* Also available with Blue, Orange or Cyan. ** Available in different size ranges.

Reagents for *in vitro* laboratory use only.

Other product sizes, combinations and customized solutions are available. Please look at www.ampliqon.com or ask for our complete product list for PCR Enzymes. For customized solutions please contact us.

Made in Denmark

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