

## Multiplex TEMPase 2x Master Mix

3 mM MgCl<sub>2</sub> final concentration

MADE IN DENMARK

Cat. No.: A260399 – SAMPLE

20 Reactions

|            |                                 |                            |
|------------|---------------------------------|----------------------------|
| -          | Multiplex TEMPase 2x Master Mix | MgCl <sub>2</sub><br>25 mM |
| ID No.     | 5500100                         | 5575801                    |
| Cap colour | Green                           | Clear                      |
| Content    | 0.5 ml                          | 1x 1.5 ml                  |

### Key Features

Multiplex PCR is a method that enables amplification of two or more amplicons simultaneously in a single reaction tube. It is widely used in genotyping and different areas of DNA testing in research, forensic and diagnostic laboratories.

Multiplex TEMPase 2x Master Mix facilitates the amplification of multiple PCR products, minimizes the need for optimization, making the development of multiplex PCR assays both fast and simple.

Multiplex TEMPase 2x Master Mix is an all-in-one 2x master mix with TEMPase Hot Start DNA polymerase, optimized buffer system, dNTPs and MgCl<sub>2</sub>. Each reaction requires 25 µl of the 2x Master Mix. Simply add primers, template and water to a total reaction volume of 50 µl to successfully carry out multiplex PCR.

TEMPase is a modified form of Ampliqon Taq DNA polymerase, which is activated by heat treatment. A chemical moiety is attached to the enzyme at the active site, which renders the enzyme inactive at room temperature. Thus, during setup and the first ramp of thermal cycling, the enzyme is not active and misprimed primers are not extended. The result is higher specificity, increased sensitivity and greater yields when compared to standard DNA polymerases, making this enzyme especially well-suited for multiplex PCR.

### Kit Components

#### Multiplex TEMPase 2x Master Mix

TEMPase Hot Start DNA polymerase, optimized buffer system, dNTPs and 6 mM mMgCl<sub>2</sub>

#### MgCl<sub>2</sub>

25 mM MgCl<sub>2</sub> in PCR grade water

#### Recommended Storage and Stability

Long term storage at -20 °C. Product expiry at -20 °C is stated on the label.

Option: Store at +4 °C for up to 6 months.

#### Quality Control

TEMPase Hot Start DNA Polymerase is tested for contaminating activities, with no traces of endonuclease activity, nicking activity or exonuclease activity.

### Protocol

This protocol serves as a guideline to ensure optimal PCR results when using Multiplex TEMPase 2x Master Mix. Optimal reaction conditions such as incubation times, temperatures, and amount of template DNA may vary and must be determined individually.

1. Thaw Master Mix and primers. **It is important to thaw the solutions completely and mix thoroughly before use to avoid localized concentrations of salts.**

**Important:** Spin vials briefly before use.

2. Prepare the reaction mix. Table 1 shows the reaction mix set-up for one reaction with a final volume of 50 µl.
3. Mix the reaction mix thoroughly and dispense appropriate volumes into reaction tubes.

**Table 1. Reaction mix and template DNA**

| Component                   | Vol./reaction*    | Final concentration*   |
|-----------------------------|-------------------|--|
| Master Mix                  | 25 µl             | 1x   |
| 25 mM MgCl <sub>2</sub>     | 0 µl (0 – 4 µl)   | 3 mM (3 – 5 mM)  |
| each forward Primer (10 µM) | 1 µl (0.5 – 5 µl) | 0.2 µM (0.1 – 1.0 µM)  |
| Each reverse primer (10 µM) | 1 µl (0.5 – 5 µl) | 0.2 µM (0.1 – 1.0 µM)  |
| PCR-grade H <sub>2</sub> O  | X µl              | -  |
| Template DNA                | X µl              | genomic DNA: 200 ng (10 – 500 ng)<br>plasmid DNA: 0.5 ng (0.1 – 1 ng)<br>bacterial DNA: 5 ng (1 – 10 ng) |
| <b>TOTAL volume</b>         | 50 µl             | -  |

\* Suggested starting conditions; theoretically used conditions in brackets

4. Add template DNA to the individual tubes containing the reaction mix.
5. Program the thermal cycler according to the manufacturer's instructions. **Each program must start with an initial heat activation step at 95°C for 15 minutes.** See table 2 for an example.  
For maximum yield and specificity, temperatures and cycling times should be optimized for each new template or primer pair.
6. Place the tubes in the thermal cycler and start the reaction.

**Table 2. Three-step PCR program**

| Cycles  | Duration of cycle  | Temperature                  |
|---------|--|------------------------------|
| 1       | 15 minutes <sup>a</sup>  | 95 °C                        |
| 30 – 40 | 20 – 30 seconds <sup>b</sup><br>50 – 90 seconds <sup>c</sup><br>50 – 90 seconds <sup>d</sup> | 95 °C<br>50 – 65 °C<br>72 °C |
| 1       | 5 minutes <sup>e</sup>   | 72 °C                        |

<sup>a</sup>. For activation of the TEMPase hot start enzyme.

<sup>b</sup>. Denaturation step: This step is the first regular cycling event and consists of heating the reaction to 95 °C for 20 – 30 seconds. It causes melting of the DNA template by disrupting the hydrogen bonds between complementary bases, yielding single-stranded DNA molecules.

<sup>c</sup>. Annealing step: The reaction temperature is lowered to 50 – 65 °C for 20 – 40 seconds allowing annealing of the primers to the single-stranded DNA template. Typically, the annealing temperature is about 3 – 5 °C below the T<sub>m</sub> (melting temperature) of the primers used.

<sup>d</sup>. Extension/elongation step: TEMPase polymerase has its optimum activity temperature at 72 °C. At this step the DNA polymerase

synthesizes a new DNA strand complementary to the DNA template strand. The extension time depends on the length of the DNA fragment to be amplified. As a rule of thumb, at its optimum temperature the DNA polymerase will polymerize a thousand bases per minute.

<sup>e</sup> Final elongation: This single step is occasionally performed at a temperature of 72 °C for 5 minutes after the last PCR cycle to ensure that any remaining single-stranded DNA is fully extended.

#### Notes:

- All primers should have the same melting temperature.
- Use equal concentrations (0.2 µM) of all primers for the initial test.
- If one product gives a much stronger band than the others, reduce the primer concentration for this target.
- If one product gives a much weaker band than the others, increase the primer concentration for this target.
- Set up reaction mixtures in an area separate from that used for DNA preparation or product analysis. Working on ice is not required.
- The MgCl<sub>2</sub> concentration in the final reaction is 3 mM with this Master Mix. In some applications, more MgCl<sub>2</sub> is required for best results. Use 25 mM MgCl<sub>2</sub> to adjust the MgCl<sub>2</sub> concentration according to table 3.

**Table 3. Additional volume (µl) of MgCl<sub>2</sub> per 50 µl reaction:**

| Final MgCl <sub>2</sub> conc. in reaction (mM) | 3.0 | 3.5 | 4.0 | 4.5 | 5.0 |
|--|-----|-----|-----|-----|-----|
| Volume of 25 mM MgCl <sub>2</sub>              | 0   | 1   | 2   | 3   | 4   |

- For longer DNA targets more DNA polymerase could be added to the PCR master mix.

## Related Products

| TEMPase Hot Start Master Mixes (500 x 50 µl reactions) *            | Cat. No.       |
|---|----------------|
| 2x Master Mix A**, 1.5 mM MgCl <sub>2</sub> final concentration     | <b>A230303</b> |
| 2x Master Mix A**BLUE, 1.5 mM MgCl <sub>2</sub> final concentration | <b>A290403</b> |

\*Master mixes available also in 1.1x variants as well as 2 mM MgCl<sub>2</sub> variants, \*\*Mix A is Ammonium Buffer based, also available as Mix C based on Combination Buffer.

| Special TEMPase Master Mixes (500 x 50 µl reactions)                | Cat. No.       |
|---|----------------|
| Multiplex 2x Master Mix, 3 mM MgCl <sub>2</sub> final concentration | <b>A260303</b> |
| GC TEMPase 2x Master Mix I – for GC-rich templates                  | <b>A331703</b> |
| GC TEMPase 2x Master Mix II – for GC-rich templates                 | <b>A332703</b> |

| Taq Master Mixes (500 x 50 µl reactions)                           | Cat. No.       |
|--|----------------|
| 2x Master Mix, 1.5 mM MgCl <sub>2</sub> final concentration        | <b>A140303</b> |
| 2x Taq OptiMix CLEAR, 1.5 mM MgCl <sub>2</sub> final concentration | <b>A370503</b> |
| 2x Master Mix RED, 1.5 mM MgCl <sub>2</sub> final concentration    | <b>A180303</b> |

| Taq DNA Polymerase (500 units) * | Cat. No.       |
|----------------------------------|----------------|
| Taq DNA Polymerase 5 U/µl        | <b>A110003</b> |
| • with 10x Ammonium Buffer       | <b>A111103</b> |

\*Available in kits including one or two buffers (Ammonium Buffer, Standard Buffer or Combination Buffer). All kits include extra 25 mM MgCl<sub>2</sub>

| Hot Start DNA Polymerase (500 units) *   | Cat. No.       |
|--|----------------|
| TEMPase Hot Start DNA Polymerase, 5 U/µl | <b>A220003</b> |
| • with 10x Ammonium Buffer               | <b>A221103</b> |

\*Available in kits including one or two buffers (Ammonium Buffer, Standard Buffer or Combination Buffer). All kits include extra 25 mM MgCl<sub>2</sub>

| Buffers for DNA polymerases *      | Cat. No.       |
|------------------------------------|----------------|
| 10x Ammonium Buffer, 3 x 1.5 ml    | <b>A301103</b> |
| 10x Standard Buffer, 3 x 1.5 ml    | <b>A302103</b> |
| 10x Combination Buffer, 3 x 1.5 ml | <b>A303103</b> |
| 5x PCR Buffer RED, 6 x 1,5 ml **   | <b>A301810</b> |
| PCR Grade Water, 6 x 5 ml          | <b>A360056</b> |

\*Ammonium Buffer, Standard Buffer and Combination Buffer are also available as Mg<sup>2+</sup> free buffers, detergent free buffers and Mg<sup>2+</sup> and detergent free buffers.  
\*\*For direct gel loading and visualisation.

Reagents for *in vitro* laboratory use only.

Other product sizes, combinations and customized solutions are available. Please look at [www.ampliqon.com](http://www.ampliqon.com) or ask for our complete product list for PCR Enzymes. For customized solutions please contact us.

**Made in Denmark**  
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